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Synthesis and Absolute Configuration of Sordidin, the Male-Produced Aggregation Pheromone of the Banana Weevil, Cosmopolites sordidus

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Abstract: The racemate as well as both the enantiomers of sordidin (1-ethyl-3,5,7-trimethyl-2,8-dioxabicyclo[3,2,1]octane, 1) were synthesized, and the natural pheromone was shown to be (1S,3R,5R,7S)-(+)-1. Copyright © 1996 Elsevier Science Ltd

The banana weevil, Cosmopolites sordidus Germar, is the major pest in all banana growing countries in the world, and its larvae feed and tunnel in the rhizomes of banana plants to destroy them. The release of a volatile aggregation pheromone by male C. sordidus was first reported by Budenberg et al. in 1993.1 Subsequently in 1995, Ducrot and his coworkers² isolated 100 μ g of the major component of the pheromone, proved its bioactivity, named it sordidin, and proposed its structure including relative stereochemistry as $(1S^*,3R^*,5R^*,7S^*)-1$ (Fig. 1) by the spectroscopic and synthetic studies. This Letter reports the synthesis of (\pm) -, (+)- and (-)-1, which enabled us to assign (1S,3R,5R,7S)-stereochemistry to the naturally occurring (+)-sordidin.

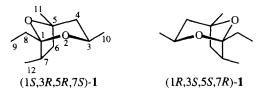


Fig. 1. Structure of sordidin.

Fig. 2 summarizes our synthesis of 1. Because Ducrot² has shown that the four diastereomers with the gross structure (\pm) -1 as well as the enantiomers of 1 are separable by GLC, the absolute configuration of sordidin must be clarified, if we synthesize 1 with known absolute configuration. The alcohol (2R)- or (2S)-5a was therefore chosen as the key intermediate to synthesize sordidin enantiomers. (\pm) -Sordidin was first synthesized in order to develop a reliable synthetic route.

Alkylation of diethyl ketone with the commercially available bromide 2 in the presence of lithium diisopropylamide (LDA) yielded 3,3 which was converted to bromoacetal 4. Lithiation of 4 with s-butyl-lithium was followed by the reaction with (\pm)-propylene oxide in the presence of boron trifluoride etherate4

Fig. 2. Synthesis of the racemate and the enantiomers of sordidin.

Reagents:(a) 1.5 eq. Et₂CO, 1.5 eq. LDA, THF (69%).— (b) 2 eq. HO(CH₂)₂OH, TsOH·H₂O, C_6H_6 (92%).— (c) 1) 1 eq. s-BuLi, THF; 2) 1.3 eq. (\pm)-propylene oxide (11); 3) 1 eq. BF₃·OEt₂ (70%).— (d) 1.5 eq. TBSCl, 3 eq. imidazole, cat. DMAP, DMF (99%).— (e) 1.5 eq. MCPBA, 5 eq. NaHCO₃, CH₂Cl₂ (75%).— (f) 8 eq. LiAlH₄, THF (69%).— (g) 1.5 eq. TsOH·H₂O, CH₂Cl₂; SiO₂ chromatog. [40% of a mixture of (\pm)-1 and (\pm)-10].— (h) prep. GLC (PEG 20M, 6 mm i.d. x 2.5 m).— (i) 1) 1 eq. s-BuLi, THF; 2) 1 eq. BF₃·OEt₂ (70%).— (j) 1) Ph₃P, PhCO₂H, EtO₂CN=NCO₂Et, THF 2) NaOMe, MeOH (82%).

to give 5a. The hydroxy group of 5a was protected as the corresponding t-butyldimethylsilyl (TBS) ether to furnish 5b, which was epoxidized with m-chloroperbenzoic acid (MCPBA) to afford 6 as a stereoisomeric mixture. Reduction of 6 with lithium aluminum hydride yielded the desired 1,3-diol 7 accompanied with the 1,4-diol 8. These were separable by silica gel chromatography, and 7 was treated with 1.5 eq. of p-toluenesulfonic acid monohydrate in dichloromethane for 4 h at room temperature. Although there existed in the reaction mixture the four stereoisomers of (\pm) -1 at the initial stage, the material isolated after 4 h was a mixture of the two acetals $[(\pm)$ -1 and (\pm) -10] and the tetrahydrofuran compound 9.5 This mixture could be separated by silica gel chromatography, and the acetals were further separated by preparative GLC6 to give (\pm) -17 and (\pm) -10.8 The spectral properties of (\pm) -1 were identical with those reported for it by the French

group.² The overall yield of the acetal mixture $[(\pm)-1+(\pm)-10]$ was 18% based on 2 (seven steps). The field evaluation of (\pm) -sordidin (1) was carried out in Venezuela. (\pm) -Sordidin attracted the banana weevils when admixed with banana plant tissue, although (\pm) -1 alone did not work. It thus works only when banana odours are present.⁹

(1R,3S,5S,7R)-(-)-Sordidin (1) and its stereoisomer (+)-10 were then synthesized via (2S)-5a by employing (S)-propylene oxide (11) and 4 as the intermediates. Mitsunobu inversion of (2S)-5a afforded (2R)-5a, which was converted to (1S,3R,5R,7S)-(+)-sordidin (1), $[\alpha]_D^{21} = +26^\circ$ (Et₂O), and its stereoisomer (-)-10, $[\alpha]_D^{21} = -7.8^\circ$ (Et₂O).¹⁰ The enantiomeric purity of (-)-1 and that of (+)-1 were estimated by their GLC analysis on a column coated with permethylated β-cyclodextrin (T. Hasegawa Co.), and found to be 95 and 92% e.e., respectively.⁶ Our synthetic enantiomers of 1 were then compared with the natural pheromone by GLC analysis (Cyclodex B column) in France, and (+)-sordidin coincided with the natural product. The absolute configuration of natural sordidin is therefore 1S,3R,5R,7S.

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References And Notes

- † Research fellow on leave from Earth Chemical Co. (1994-1996).
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- 2. Beauhaire, J.; Ducrot, P.-H.; Malosse, C.; Rochat, D.; Ndiege, I. O.; Otieno, D. O. Tetrahedron Lett. 1995, 36, 1043-1046.
- All the new compounds were characterized by spectroscopic (IR and NMR) and elemental (combustion or HRMS) analysis.
- 4. Eis, M. J.; Wrobel, J. E.; Ganem, B. J. Am. Chem. Soc. 1984, 106, 3693-3694.
- 5. The stereoisomers A of sordidin must be unstable due to the severe 1,3-diaxial interaction of the substituents of the 1,3-dioxacyclohexane ring. After protonation to give B, B generates the carbocation C, which gives the rearranged compound. Its most probable structure is 9.

Properties of 9: IR v_{max} (film) 1715 (s, C=O), 1360 (s), 995 (m), 740 (m) cm⁻¹; ¹H NMR (270MHz, CDCl₃) 0.93(1.5 H, d, J = 6.9 Hz), 0.94 (1.5H, J = 6.3 Hz), 0.96 (3H, t, J = 7.3 Hz), 1.24 (1.5H, s), 1.34 (1.5H, s), 1.30-1.65 (3H, m), 2.00 (0.5H, dd, J = 7.6, 12.9 Hz), 2.16 (0.5H, dd, J = 7.6, 12.8 Hz), 2.20 (1.5H, s), 2.21 (1.5H, s), 2.27 (0.5H, m), 2.41(0.5H, m), 2.53(0.5H, d, J = 14.2 Hz), 2.63 (0.5H, d, J = 14.5 Hz), 2.70 (0.5 H, d, J = 14.2 Hz), 2.77 (0.5H, d, J = 14.5 Hz), 3.73-3.87 (1H, m); ¹³C NMR (22.4 MHz, CDCl₃) δ

- 10.9, 14.6, 14.7, 23.8, 23.9, 26.7, 28.8, 31.8, 35.4, 36.0, 45.2, 45.7, 54.6, 56.4, 79.6, 79.8, 82.0, 82.6, 207.6, 208.1 (diastereomeric mixture); GC-MS (70 eV) i) The isomer with a shorter Rt :m/z 43 (100), 55 (10), 69 (10), 83 (8), 95 (12), 97 (9), 111 (12), 155 (9), 169 (1), 184 (M+, < 0.05), ii) The isomer with a longer Rt :m/z 43 (100), 55 (9), 69 (11), 83 (6), 95 (7), 97 (9), 111 (7), 127 (8), 169 (0.2), 184 (M+, 0.3).
- 6. GC conditions; preparative—PREPGC-TH (special GC) equipped with a PEG-20M, 10% on Uniport-HP (80-100 mesh), 100°C (constant). analysis—GC-14A equipped with a PEG-20M(0.25 mm i.d. x 60 m), 120°C (constant). chiral analysis—GC-14A equipped with a DMPBCD-TH {heptakis-(2,6-di-O-methyl-3-O-pentyl)-β-cyclodextrin}, 0.25 mm i.d. x 50 m, 70°C to 140°C (1.0°C/min).
- Properties of (\pm) -1: $n_D^{21.4}$ 1.4468; IR v_{max} (film) 2980 (s), 2940 (s), 2885 (s), 1455 (m), 1377 (s), 1200 (s), 1135 (s), 1005 (s), 945 (s) cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.98 (3H, t, J = 7.5 Hz, 9-H), 0.98 (3H, d, J = 7.0 Hz, 12-H), 1.16 (3H, d, J = 6.1 Hz, 10-H), 1.18 (1H, ddd, J = 4.6, 12.7, 1.0 Hz, 6_{exo} -H), 1.30 (3H, s, 11-H), 1.34 (1H, ddd, J = 6.1, 13.0 Hz, 4-Heq), 1.35 (1H, br dd, J = 8.9, 13.0 Hz, 4-Hax), 1.62 (1H, dq, J = 14.0, 7.5 Hz, 8-H), 1.71 (1H, dq, J = 14.0, 7.5 Hz, 8-H), 2.15 (1H, dd, J = 8.9, 12.7 Hz, 6_{endo} -H), 2.33 (1H, ddq, J = 8.9, 4.6, 7.0 Hz, 7-H), 3.95 (1H, ddq, J = 6.1, 8.9, 6.1 Hz, 3-H); ¹³C NMR (125 MHz, CDCl₃) δ 7.98 (C-9), 19.87 (C-12), 21.91 (C-10), 26.53 (C-11), 27.41 (C-8), 40.01 (C-7), 44.12 (C-4), 44.87 (C-6), 64.51 (C-3), 78.74 (C-5), 108.59 (C-1); GCMS (70 eV) m/z: 41 (24), 43 (78), 57 (100), 67 (13), 69 (9), 71 (8), 83 (17), 85 (11), 95 (78), 100 (5), 113 (16), 125 (2), 142 (9), 151 (1), 169(1), 184 (M+, 0.5); HRMS: Calc. for C₁₁H₂₀O₂ = 184.1464, Found 184.1447; bp 100-105°C(bath temp.)/110 Torr for the mixture of (±)-1 and (±)-10. When the pure (±)-1 was treated with p-toluenesulfonic acid in dichloromethane, an equilibration mixture of (±)-1 and (±)-10 (48 : 52) was obtained.

The assignment of underlined signals is different from that reported.² We confirmed this assignment by the NMR experiments (¹H-¹H cosy, ¹H-¹H noesy, ¹H-¹³C cosy and HMBC).

- 8. Properties of (\pm) -10: $n_D^{21.4}$ 1.4446: IRv_{max} (film) 2980 (s), 2940 (s), 2885 (s), 1455 (m), 1377 (s), 1200 (s), 1135 (s), 1005 (s), 945 (s) cm⁻¹: ¹H NMR (500 MHz, CDCl₃) δ 0.95 (3H, t, J = 7.5 Hz, 9-H), 1.08 (3H, d, J = 7.5 Hz, 12-H), 1.18 (3H, s, J = 6.1 Hz, 10-H), 1.32 (3H, s, 11-H), 1.39 (1H, dd, J = 4.5, 13.0 Hz, 4-Heq), 1.44 (3H, br dd, J = 10.5, 13.0 Hz, 4-Hax), 1.48 (1H, dd, J = 6.5, 12.5 Hz, 6_{endo} -H), 1.56 (1H, dq, J = 14.5, 7.5 Hz, 8-H), 1.72 (1H, dq, J = 14.5, 7.5 Hz, 8-H), 1.99 (1H, ddd, J = 1.5, 12.5, 12.5 Hz, 6_{exo} -H), 2.25 (1H, ddq, J = 6.5, 12.5, 7.5 Hz, 7-H), 4.08 (1H, ddq, J = 4.5, 10.5, 6.1 Hz, 3-H); ¹³C NMR (125 MHz, CDCl₃) δ 7.87 (C-9), 12.72 (C-12), 22.20 (C-10), 26.45 (C-11), 29.04 (C-8), 40.57 (C-7), 42.38 (C-4), 44.54 (C-6), 65.57 (C-3), 78.68 (C-5), 107.55 (C-1); GCMS (70 eV) m/z: 41 (24), 43 (78), 57 (100), 67 (13), 69 (9), 71 (8), 83 (17), 85 (11), 95 (78), 100 (5), 113 (16), 125 (2), 142 (9), 151 (0), 169 (1), 184 (M+, 0.5); HRMS: Calc. for $C_{11}H_{20}O_2$ = 184.1464, Found 184.1453.
- 9. Ditails of the bioassay of (±)-1 will be published separately by Prof. K. Jaffe in J. Chem. Ecol.
- 10. Properties of the optically active products: (1) (+)- $\mathbf{1}$ — $n_D^{21.6}$ 1.4471; $[\alpha]_D^{21}$ +26° (c = 0.48, Et₂O), (2) (-)- $\mathbf{1}$ — $n_D^{22.3}$ 1.4457; $[\alpha]_D^{21}$ -26° (c = 0.59, Et₂O), (3) (+)- $\mathbf{10}$ — $n_D^{22.3}$ 1.4449; $[\alpha]_D^{21}$ +7.9° (c = 0.67, Et₂O), (4) (-)- $\mathbf{10}$ — $n_D^{21.6}$ 1.4443; $[\alpha]_D^{21}$ -7.8° (c = 0.48, Et₂O).

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